

EXHIBIT H

Guelcher PCT-168 Documents

PCT-167 BSC

PCT-168 Ethicon SEM Testing will also be conducted on select FTIR samples.

in vitro testing

42 samples			42 samples			36 samples			15 samples		
PCT-167 Advantage Sample ID	Sample No.	Planned Exposure to Oxidizing Solution (weeks)	PCT-167 Lynx Sample ID	Sample No.	Planned Exposure to Oxidizing Solution (weeks)	PCT-168 TVT Sample ID	Sample No.	Planned Exposure to Oxidizing Solution (weeks)	PP STD Sample ID	Sample No.	Planned Exposure to Oxidizing Solution (weeks)
FTIR	1	0	FTIR	1	0	FTIR	1	0	FTIR	1	0
FTIR	2	0	FTIR	2	0	FTIR	2	0	FTIR	2	0
XPS	3	0	XPS	3	0	XPS	3	0	XPS	3	0
FTIR	4	1	FTIR	4	1	FTIR	4	1	FTIR	4	1
FTIR	5	1	FTIR	5	1	FTIR	5	1	FTIR	5	1
FTIR	6	1	FTIR	6	1	FTIR	6	1	XPS	6	1
XPS	7	1	XPS	7	1	XPS	7	1	XPS	7	3
XPS	8	1	XPS	8	1	XPS	8	1	FTIR	8	3
XPS	9	1	XPS	9	1	XPS	9	1	FTIR	9	3
FTIR	10	2	FTIR	10	2	FTIR	10	2	XPS	10	4
FTIR	11	2	FTIR	11	2	FTIR	11	2	FTIR	11	4
FTIR	12	2	FTIR	12	2	FTIR	12	2	FTIR	12	4
XPS	13	2	XPS	13	2	XPS	13	2	FTIR	13	2
XPS	14	2	XPS	14	2	XPS	14	2	FTIR	14	2
XPS	15	2	XPS	15	2	XPS	15	2	XPS	15	2
FTIR	16	3	FTIR	16	3	FTIR	16	3			
FTIR	17	3	FTIR	17	3	FTIR	17	3			
FTIR	18	3	FTIR	18	3	FTIR	18	3			
XPS	19	3	XPS	19	3	XPS	19	3			
XPS	20	3	XPS	20	3	XPS	20	3			
XPS	21	3	XPS	21	3	XPS	21	3			
FTIR	22	4	FTIR	22	4	FTIR	22	4			
FTIR	23	4	FTIR	23	4	FTIR	23	4			
FTIR	24	4	FTIR	24	4	FTIR	24	4			
XPS	25	4	XPS	25	4	XPS	25	4			
XPS	26	4	XPS	26	4	XPS	26	4			
XPS	27	4	XPS	27	4	XPS	27	4			
FTIR	28	5	FTIR	28	5	FTIR	28	5			
FTIR	29	5	FTIR	29	5	FTIR	29	5			
FTIR	30	5	FTIR	30	5	FTIR	30	5			
XPS	31	5	XPS	31	5	XPS	31	5			
XPS	32	5	XPS	32	5	XPS	32	5			
XPS	33	5	XPS	33	5	XPS	33	5			
FTIR	34	6	FTIR	34	6	FTIR	34	6			
FTIR	35	6	FTIR	35	6	FTIR	35	6			
FTIR	36	6	FTIR	36	6	XPS	36	6			
XPS	37	6	XPS	37	6						
XPS	38	6	XPS	38	6						
XPS	39	6	XPS	39	6						
FTIR	40	TBD	FTIR	40	TBD						
FTIR	41	TBD	FTIR	41	TBD						
XPS	42	TBD	XPS	42	TBD						

Procedure

1. Prepare oxidative media using the separate protocol.
2. Obtain all mesh samples by cutting from the respective exemplar mesh (each sample is approx 1/4in x 1/2 inch)
3. Place each mesh sample in individually labeled glass vials and photo document
4. Place two layers of glass beads on top of the mesh in each vial to keep the mesh submerged in the media. Use a perforated teflon disc for the peiler samples.
5. Place ~5ml of oxidative media in each vial.
6. Insert all vials in the vial holder.
7. Place the vial container on the rotator in the incubator. Set the rotator to a medium setting. Set the incubator at 37°C.
8. Remove samples from the incubator as shown in the above testing schedule.
9. Using gloves and tweezers, remove the appropriate mesh sample from its vial and rinse the mesh sample with deionized (DI) water to rinse off residual oxidative media.
10. Blot dry the mesh sample using Kim Wipes.
11. Dry the empty vial using compressed air.
12. Place the dry mesh sample back into the dry glass vial.
13. Test the dry mesh samples using the method specified for each respective sample. Retain each mesh sample in its vial after testing and place in storage.
14. Run SEM analysis on samples as needed. Mark the samples used for SEM testing with an additional label on the vial.

Notes:

Samples initially placed into oxidizing medium on Friday, Sept. 19
 Oxidizing medium replaced on Wednesday, Sept. 24
 Week 1 samples removed from oxidizing medium on Friday, Sept. 26
 Oxidizing medium replaced on Wednesday, Oct. 1
 Week 2 samples removed from oxidizing medium on Friday, Oct. 3
 Oxidizing medium replaced on Wednesday, Oct. 8
 Week 3 samples removed from oxidizing medium on Friday, Oct. 10
 Oxidizing medium replaced on Tuesday, Oct. 14
 Week 4 samples removed from oxidizing medium on Thursday, Oct. 16
 Oxidizing medium replaced on Thursday, Oct. 23
 Week 5 samples removed from oxidizing medium on Friday, Oct. 24
 Oxidizing medium replaced on Wednesday, Oct. 29
 Week 6 samples removed from oxidizing medium on Friday, Oct. 31
 Final samples removed from oxidizing medium on Saturday, Dec. 6

Guelcher Lab
Oxidative Media Preparation

Standard Operating Procedure ###**Principle:**

Prepare oxidative degradation media for *in vitro* degradation studies. This media will contain 20 wt% hydrogen peroxide and 0.1M cobalt chloride.

Before starting:

- Read and understand the MSDS of the reagents listed below
- Personal Protective and Safety Equipment required:
 - Disposable nitrile gloves
 - Heavy duty gloves
 - Hood
 - Appropriate attire according to the Chemical Hygiene Plan (shoes, labcoat, goggles, etc.)

Reagents:

- 30 wt% hydrogen peroxide (Sigma Aldrich)
- Cobalt chloride hexahydrate (Fisher)
- Water

Materials and Equipment:

- 1 beaker (size depends on batch size)
- Glass bottle
- Foil
- Stir plate
- Stir bar

Procedure (All listed values are for a 1L batch size)

1. Make sure all glassware is washed and dried prior to use
2. Dissolve cobalt chloride in water (23.78 g CoCl_2 hexahydrate in 333 mL water) in a glass beaker
3. In the glass bottle, measure out required amount of 30 wt% hydrogen peroxide (667 mL)
4. Add CoCl_2 solution to hydrogen peroxide slowly while stirring
5. Cover glass bottle with foil and store in the refrigerator

Notes:

- Addition of CoCl_2 to H_2O_2 can produce heat and gas. Make this solution in the hood.
- $\text{CoCl}_2(\text{s})$ will react vigorously with H_2O_2 , so make sure CoCl_2 is fully dissolved

Clean-up:

1. Collect all glass waste (pipettes, vials, or broken glass) and dispose in the broken glass container (box)
2. Collect all sharps and dispose in the sharps waste container (red box)
3. Oxidative media waste contains heavy metals and must be collected and disposed of properly
4. Clean glassware:
 - a. Wash with soap and water
 - b. Rinse with acetone and dry in the oven